

## **Guidelines for the Blood Transfusion Services**

### **Chapter 14: Guidelines for the use of DNA/PCR techniques in Blood Establishments**

<http://transfusionguidelines.org/red-book/chapter-14-guidelines-for-the-use-of-dna-pcr-techniques-in-blood-establishments>

## **Chapter 14:**

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### **Guidelines for the use of DNA/PCR techniques in Blood Establishments**

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#### **14.1: Safety precautions**

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All human cells should be treated as potentially infectious. Materials should be handled and discarded according to in-house documented procedures for potentially infectious biological materials.

Operators working with ultraviolet (UV) light should wear opaque gloves and a UV protective visor, appropriate to the wavelength emitted. Exposure should be kept to a minimum.

Alternatives to ethidium bromide for gel based detection of PCR products should be used where possible.

#### **14.2: Avoidance of contamination**

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DNA should be purified by a standard method that has been reported to the scientific literature and validated in the laboratory. DNA should be suitably stored to protect the integrity of the material.

During the preparation of genomic DNA, great care should be taken to avoid contamination from any other source of DNA. Pre-polymerase chain reaction (PCR) and post-PCR procedures should be undertaken in separate areas and using separate laboratory coats in each area. The laboratory should have documented procedures which have been constructed to eliminate potential causes of contamination, including training of the operator. If contamination does occur, all procedures should be reviewed and appropriate corrective action taken. Proposed change to procedures should be validated prior to their introduction.

In order to avoid contamination, the use of separate working stations or clearly defined work areas is beneficial for each stage of the PCR process. For example:

- One to prepare reagents. This is particularly important to avoid contamination of primers.
- One dedicated to pre-PCR manipulation (e.g. DNA isolation). A Class II laminar flow cabinet should avoid contamination of the sample with DNA from the operator.
- One dedicated to setting up PCRs.
- One for manipulation of PCR-amplified DNA. PCR-amplified products should be kept away from areas used for pre-amplification manipulation and reagent preparation.

Each working station should be adequately and independently equipped. However, the use of such working stations should not absolve the laboratory from ensuring procedures are constructed to eliminate contamination.

Examples of measures which will help to minimise contamination include:

- the use of new sterilised, disposable plastic tubes or glassware for handling DNA
- the use of freshly prepared and sterilised materials and reagents when making up solutions for DNA samples, particularly dH<sub>2</sub>O and buffers
- aliquoting reagents in small amounts to minimise the number of repeat samplings
- the changing of gloves and coats when moving between the areas dedicated for pre- and post-PCR manipulations
- the use of positive displacement dedicated pipettes or plugged tips to carry out PCR preparations
- routine wipe-tests of pre-amplification work areas should be performed. If an amplified product is detected, the area must be cleaned to eliminate the contamination, re-tested and measures taken to prevent future contamination
- reagents used for amplification must not be exposed to post-amplification work areas.

### 14.3: Working practices

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- DNA should be as intact as possible.
- An archival record (e.g. photograph or electronic image) of each post-PCR run should be retained.
- The performance of non-commercial kit based probes and primers must be fully validated and characterised before they are put into use.
- Reagents (e.g. chemicals, enzymes) must be stored and utilised under conditions recommended by the manufacturer, including, for example, storage temperature, test temperature, shelf life, diluent buffer and concentration for use.
- Each lot of reagents must be tested before use in routine typing.
- For reagents and kits, the source, lot number, expiration date and storage conditions should be documented.
- Users should have procedures to ensure that periodic checks of probes and primers are carried out to detect their deteriorating performance or contamination.
- Thermal cyclers should be serviced at least annually according to the manufacturer's recommendations and a temperature calibration should be performed. A record of the service and calibration checks should be maintained.
- When using non-commercial kit testing methods laboratories should regularly check their primer sequences for newly discovered single nucleotide polymorphisms. This can be done via the website for the National Genetics References Laboratories ([ngri.org.uk](http://ngri.org.uk)).
- Software used for analysis of results must be validated before use and updated regularly with appropriate allele sequences.